

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/319410310>

Sparse and Burst Spiking in Artificial Neural Networks inspired by Synaptic Retrograde Signaling

Article in *Information Sciences* · August 2017

DOI: 10.1016/j.ins.2017.08.073

CITATIONS

0

READS

29

2 authors:



Faramarz Faghihi

Shahid Beheshti University of Medical Sciences

12 PUBLICATIONS 17 CITATIONS

[SEE PROFILE](#)



Ahmed A Moustafa

Western Sydney University

141 PUBLICATIONS 2,176 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Bioinspired artificial neural networks [View project](#)



Computational neuroscience models [View project](#)

All content following this page was uploaded by [Faramarz Faghihi](#) on 01 September 2017.

The user has requested enhancement of the downloaded file.

AUTHOR QUERY FORM

 <p>ELSEVIER</p>	<p>Journal: INS</p> <p>Article Number: 13078</p>	<p>Please e-mail your responses and any corrections to:</p> <p>E-mail: correctionsaptara@elsevier.com</p>
--	--	---

Dear Author,

Please check your proof carefully and mark all corrections at the appropriate place in the proof (e.g., by using on-screen annotation in the PDF file) or compile them in a separate list. Note: if you opt to annotate the file with software other than Adobe Reader then please also highlight the appropriate place in the PDF file. To ensure fast publication of your paper please return your corrections within 48 hours.

Your article is registered as a regular item and is being processed for inclusion in a regular issue of the journal. If this is NOT correct and your article belongs to a Special Issue/Collection please contact s.sandacoumar@elsevier.com immediately prior to returning your corrections.

For correction or revision of any artwork, please consult <http://www.elsevier.com/artworkinstructions>

Any queries or remarks that have arisen during the processing of your manuscript are listed below and highlighted by flags in the proof. Click on the '[Q](#)' link to go to the location in the proof.

Location in article	Query / Remark: click on the Q link to go Please insert your reply or correction at the corresponding line in the proof		
Q1	<p>AU: The author names have been tagged as given names and surnames (surnames are highlighted in teal color). Please confirm if they have been identified correctly.</p> <table border="1" data-bbox="492 1268 1118 1373"><tr><td data-bbox="492 1268 1052 1373">Please check this box or indicate your approval if you have no corrections to make to the PDF file</td><td data-bbox="1052 1268 1118 1373"></td></tr></table>	Please check this box or indicate your approval if you have no corrections to make to the PDF file	
Please check this box or indicate your approval if you have no corrections to make to the PDF file			

Thank you for your assistance.



Contents lists available at ScienceDirect

Information Sciences

journal homepage: www.elsevier.com/locate/ins

Sparse and burst spiking in artificial neural networks inspired by synaptic retrograde signaling

Faramarz Faghihi^a, Ahmed A. Moustafa^{b,*}^aInstitute for Cognitive and Brain studies, Shahid Beheshti University, Tehran, Iran^bSchool of Social Sciences and Psychology & Marcs Institute for Brain and Behaviour, University of Western Sydney, Sydney, New South Wales, Australia

ARTICLE INFO

Article history:

Received 31 January 2017
Revised 26 July 2017
Accepted 20 August 2017
Available online xxx

Keywords:

Retrograde signalling
Neurotransmitter release
Spiking neural networks
Closed loop

ABSTRACT

The bursting of action potential and sparse activity are ubiquitously observed in the brain. Although the functions of these activity modes remain to be understood, it is expected that they play a critical role in information processing. In addition, the functional role of retrograde signalling in neural systems is under intensive research. Therefore, we propose a bio-inspired neural network that is capable of demonstrating these activity modes as well as shifting themselves from normal to bursting or sparse modes by changing model parameter values. Accordingly, we model diffused retrograde signalling with different activity patterns in dendrites and presynaptic neurons. Using in a three-layered spiking neural network, simulation studies are conducted using different conditions and parameter values to find factors underlying the change in firing rate of output neurons. Our findings propose the application of retrograde signalling as a known synaptic mechanism for the development of artificial neural systems to encode environmental information by different spiking modes.

© 2017 Elsevier Inc. All rights reserved.

1. Introduction

Neurons communicate with other neurons by transforming synaptic input patterns into output spike trains. This mode of communication strongly depends on the properties of voltage-gated conductance in neuronal membranes. Biological neural systems are equipped with complex molecular and synaptic mechanisms [15,45]. Specifically, synaptic mechanisms play a key role in memory and learning processes that take place across different time scales [40,36]. Although understanding the function of neural systems at different time scales is a challenge for modern neurosciences, theoretical and computational studies may help understand and interpret experimental findings by shedding light on how single neurons and neural populations encode, store and retrieve information [27]. Moreover, to develop artificial architectures that show animal-like behaviours, we must understand the principles of information processing in the neural systems and how they trigger behaviours.

Bursting is a firing mode of neurons which is characterized by high frequency spikes, followed by a period of relative silence [7].

Neurons in a variety of biological neural systems exhibit correlated activity. Understanding how input correlations are processed and transmitted from a layer of neurons to the next neural layer has been studied. Recently, it has been shown

* Corresponding author.

E-mail address: ahmedhalimo@gmail.com (A.A. Moustafa).

<http://dx.doi.org/10.1016/j.ins.2017.08.073>

0020-0255/© 2017 Elsevier Inc. All rights reserved.

15 that neural bursting can be generated intrinsically in neurons themselves or as a result of stimulation by network activity
16 [24] that plays a central role in enhancing output correlations of a neural layer. Another observed firing mode of neurons
17 is sparse spiking –low firing rate– which is observed in some brain regions such as the dentate gyrus in the hippocampus
18 [11] or auditory cortex [26].

19 Activity in cortical networks is heterogeneous, sparse and often precisely timed. The functional significance of sparseness
20 and precise spike timing has been under intense debates. Some studies have provided an account for the developmental
21 and synaptic mechanisms that shape neuronal discharge sparse patterns in cortical activities [40].

22 It is believed that feedforward or feedback inhibitory circuits in neural networks plays a role in controlling sparse activity
23 of cortical neural networks [23,32,10]. These inhibition mechanisms play a key role in memory processes in insects [13].
24 In addition, the homeostatic feedback mechanism in synapses is one of the stabilizing mechanisms in which diffusion of
25 retrograde messengers from postsynaptic to presynaptic neurons as a consequence of action potential reach axonal terminals
26 [35]. These mechanisms equip neurons and neuronal circuits to sense how active they are and to adjust their firing to keep
27 this activity within some target range [34]. In addition, biophysical features of neurons including factors that determine
28 excitability of neurons are other mechanisms that impact sparse and burst spiking neural activities [8].

29 Although neurons mainly communicate via generating action potentials in short time scales, this is not the only way
30 they transmit information to other neurons in order to induce changes including the stimulation of postsynaptic neurons.
31 In response to stimulation from presynaptic neurons, neurons may generate chemicals of different types that are diffused
32 into presynaptic neurons [15,25]. These chemicals may induce some changes in the cellular and molecular activities that
33 eventually result in a change of electrical activity of neurons at different time scales [4]. The importance of some of these
34 non-synaptic mechanisms in healthy and abnormal neural functions is relatively well-known [14].

35 Retrograde signalling in neurons plays a role in information processing in healthy brains while abnormal retrograde
36 signalling (low or high levels of retrograde messengers) may lead to brain disorders [28,29,38]. Nitric oxide is one of the
37 well studied fast diffused retrograde messengers, and its abnormal levels are shown to lead to psychiatric disorders [1].
38 Further, repetitive synaptic activity can induce persistent increase or decrease of synaptic efficacy. Retrograde signalling as
39 diffused chemicals from postsynaptic neuron to presynaptic neurons is essential for the induction of long-term potentiation
40 (LTP) or long-term depression (LTD) [30].

41 One of the changes that such retrograde signals can induce in presynaptic neurons is an increase or decrease in the
42 probability of neurotransmitter release from presynaptic neurons [9]. The importance of neurotransmitter release probability
43 as a consequence of presynaptic activities of different neuron types and its role in encoding efficiency of neural systems has
44 been studied using computational modelling [9]. Regarding the initial probability of neurotransmitter release in different
45 neurons, synapses are categorized as filters of information [2]. The initial value of release probability is believed to be
46 justified by diffused retrograde signalling from postsynaptic into presynaptic neurons. These mechanisms, in combination
47 with synaptic and intrinsic excitability, have been studied to determine the overall effect on the activity of neural population
48 to encode and store the information from different sources of stimulations [37].

49 Biological neurons have complex and diverse shapes and sizes, and electrophysiological features. Therefore, to simulate
50 their function in a network, it is necessary to simplify biological features of neurons. Artificial Neural Networks incorporate
51 basic information processing from biological neurons and their biophysical features that process sensory input processing
52 and generate action potential to transfer information to the networks [21]. Models that simplify neuronal electrophysiology
53 are computationally efficient but they are generally very abstract to be used in biologically realistic simulations [20].

54 Some simulation techniques of neuron models that consider biophysical feature of neurons in details are computationally
55 expensive and unsuitable for the simulation of large aggregate of neurons, such as network simulations [20].

56 Bio-inspired neural networks developed with different architectures (e.g., multilayer perceptron classifiers [19]) have
57 many industrial applications including optimization [41], dealing with large scale datasets [6], and image processing (e.g.,
58 handling the human pose recovery problem [17,18]). For this purpose, retrieving of images with sparse coding by artificial
59 neurons has been effectively used.

60 One of such bio-inspired neural systems is deep neural networks which have been used to solve image processing prob-
61 lems by high-dimensional sparse representation [43]. In this line of research, to obtain an appropriate description of images,
62 multimodal features have been considered for describing images. For this purpose, recently, a novel deep multimodal dis-
63 tance metric learning combines these multimodal features [44].

64 In this study, we have developed a three-layer feedforward neural network model using a hypothetical mechanism of
65 changing the probability of neurotransmitter release that is induced by diffused retrograde messengers from postsynaptic
66 neurons. Synapses that show production and diffusion of retrograde messengers from a postsynaptic neuron in response
67 to presynaptic stimulation are considered as a closed loop [22]. In this study, we have modelled controlling of neuro-
68 transmitter release by a closed loop in presynaptic and postsynaptic cellular machinery. The neural system is able to decrease
69 or increase the probability of neurotransmitter release in response to different levels of stimulation. The role of different
70 model parameter values is examined using simulation studies to investigate neural system's dynamic activation in differ-
71 ent conditions of stimulation. Such feedforward neural system as divergence-convergence architecture is found in many
72 sensory-perception loops in animals' brains. Two well-known examples are *Drosophila* olfactory system and information pro-
73 cessing in hippocampus. In the insect brain, activated neurons in Antennal Lobe (about 150 neurons) transmit information
74 to Mushroom Body (memory center of insects that includes about 2500 neurons) [16]. The activated neurons in Mushroom
75 Body project to few Output Neurons that elicit avoidance behavior. In the rat hippocampus, about 200,000 neurons in the

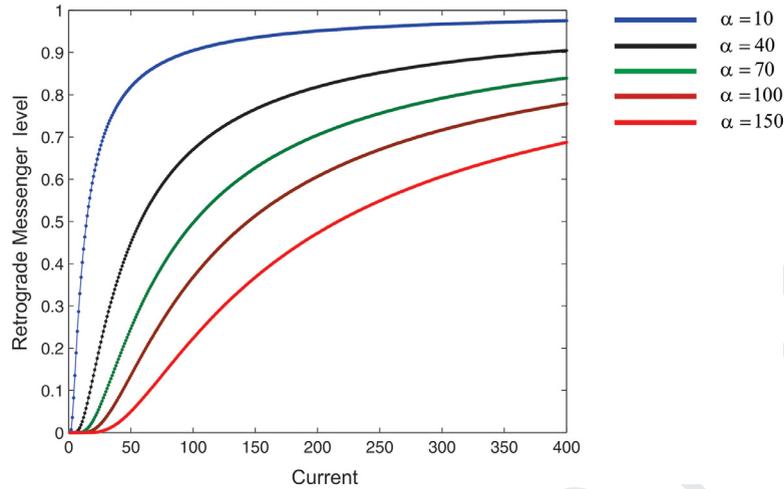


Fig. 2. The production of retrograde messenger in post-synaptic neuron (third layer) induced by total current from pre-synaptic neurons (second layer). Different model parameters (α values) determine the level of a produced retrograde signal at a given current.

98 The current into L_2 neurons and L_3 neurons are modeled as Eq. (2)

$$\dot{I} = -I(t)/\tau_I + \sum_{t_p} \delta(t - t_p); \quad \tau_I = 30 \text{ ms} \quad (2)$$

99 where $I(t)$ denotes current into a postsynaptic neuron in time bin 't'. where $\delta(t - t_p)$ is the Dirac function that step-increases
100 the variable I .

101 $\sum_{t_p} \delta(t - t_p)$ is the sum of the received spikes as input in a given time bin. The input intensity depends on both firing
102 rate of pre-synaptic neurons, connectivity rate of layers and probability of neurotransmitter release in each pre-synaptic
103 neuron. t_p is the time bin in which a spike is received. The total amount of current that a neuron in L_3 receives from
104 connected neurons of L_2 at the end of each time bin (t) triggers the post-synaptic neuron to produce retrograde messenger
105 (Eq. 3). α determines the level of produced and received retrograde messenger for a given level of current into post-synaptic
106 neuron (Fig. 2).

$$RM(t) = \exp(-\alpha/I^{tot}) \quad (3)$$

107 where I^{tot} denotes total current into postsynaptic neurons at the beginning of time bin 't'.

108 In this study, in all simulations $\alpha = 120$ is used.

109 To model the effect of retrograde messenger produced in post-synaptic neurons (L_3) and received by pre-synaptic neu-
110 rons (L_2) we considered different retrograde messengers with slow to fast effect on neurotransmitter release machinery by
111 τ_{RM} values (Eq. 4).

$$RM_{ef}(t) = RM \frac{t}{\tau_{RM}} \exp\left(\frac{-t}{\tau_{RM}}\right) ; \tau_{RM} = [10 \ 200] \quad (4)$$

112 where RM in the right hand of Eq. 4 is the total amount of received retrograde messenger by a L_2 neurons at the end of
113 each time bin.

114 Low τ_{RM} values induce fast effect on release machinery and fast decay of effect as well. High τ_{RM} values induce slow
115 effect and slow decay of activity of release machinery (Fig. 3). Fig. 3 shows the increase and decay of retrograde messenger's
116 effect on presynaptic neuron triggered at time bin equal to 1. In the simulations where RM may produce in a sequence of
117 time bins, RM_{ef} may increase over time.

118 2.2. Synapse dynamics

119 The spiking activity of each neuron in L_2 may result in neurotransmitter release into the cleft that consequently leads to
120 influx of current into the postsynaptic site (L_3) (Eq. 2).

121 The neurons in L_2 has an initial release probability that can be decreased or increased through neuronal activity and dif-
122 ferent stimuli representation conditions that occur by the synaptic dynamics model developed in this study. In this model,
123 it is assumed that presynaptic neurons are equipped with release machinery with an activity threshold (θ) that determines
124 either decrease or increase in neurotransmitter release probability that is calculated at the end of each time bin. Two pa-
125 rameters, K and M , are used in order to modify the decrease or increase of the release probability, respectively. If the level
126 of induced activity of release machinery in each time bin gets a lower value compared to the threshold (θ) (Eq. 5) then the
127 system increases the previous release probability (Eq. 6); otherwise, it leads to the decrease in the neurotransmitter release

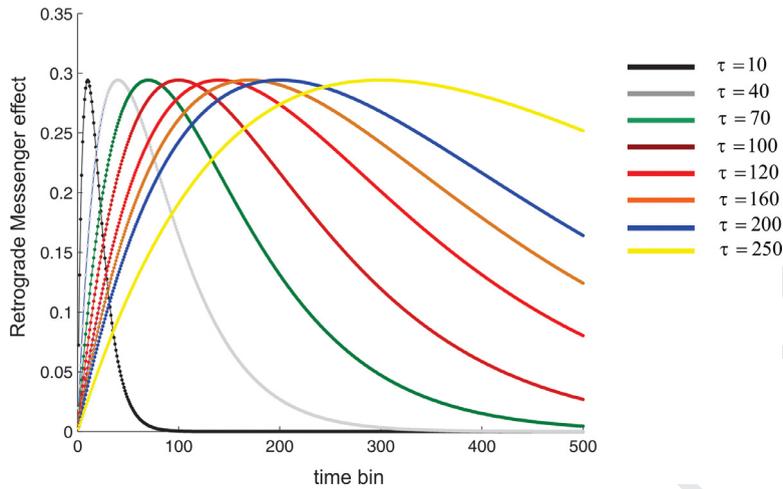


Fig. 3. The effects of diffused retrograde signalling from a post-synaptic neuron on activity of pre-synaptic neurons. This activity acts as a part of the 'controller' to determine probability of neurotransmitter release into a post-synaptic neuron. Different parameter values (τ) determine the biophysical features of retrograde signal like diffusion into a pre-synaptic neuron and induced activity inside a pre-synaptic neuron. Higher τ values demonstrate retrograde signalling that are slowly diffused and act on pre-synaptic neurons.

Table 2
Summary of notations.

L_{1-3}	layers in feedforward neural network
r_{1-2}	connectivity rates of layers
$I(t)$	current into postsynaptic neuron
δ	Dirac function
t_p	time of receiving a spike
$\sum I$	total current into I&F neuron
$RM(t)$	retrograde messenger concentration
$RM_{ef}(t)$	induced changes in presynaptic neuron by RM
α	parameter of retrograde messenger production
τ_{RM}	parameter of diffusion of retrograde messenger
θ	threshold of RM_{ef} to increase or decrease P_{rel}
K, M	parameters of release probability
$E(t)$	error function
P_{rel}	release probability

128 probability expressed as Eq. 7. The difference between release machinery and the threshold can be expressed as 'error (E)'.
 129 For the sake of simplicity in implementing the model, release probability is calculated and updated as a difference equation
 130 at the beginning of each time bin.

$$E(t) = RM_{ef}(t) - \theta; \theta = [0.5 \ 10] \quad (5)$$

131

$$\text{If } E(t) < 0 \quad (6)$$

$$P_{rel}^{t+1} = P_{rel}^t + M.(E(t))^2$$

132

$$\text{If } E(t) > 0 \quad (7)$$

$$P_{rel}^{t+1} = P_{rel}^t - K.(E(t))^2$$

133 K and M determine the rate of increase or decrease in release probability in the successive time bin. Initial release proba-
 134 bility in all simulations is set to 0.95.

135 Using the changes of release probability expressed as Eqs. 5 to 7 some simulations are run in order to study the role of
 136 model parameters in different conditions. Summary of notations defined and used in the model is resented in Table 2.

137 2.3. Robustness of the system

138 The developed neural system demonstrates both sparse and burst spiking. To study how model parameter values ensure
 139 burst spiking activities in high level of stimulation (as L_1 neurons firing rate equal to 0.7), K and M values are set to 0.2
 140 and 0.1, respectively. Then the stimulation level is lowered and the optimal values of θ and τ_{RM} are investigated to keep the
 141 L_3 firing rate at high levels. To obtain sparse spiking activities in high level of stimulation (as L_1 neurons firing rate equal to

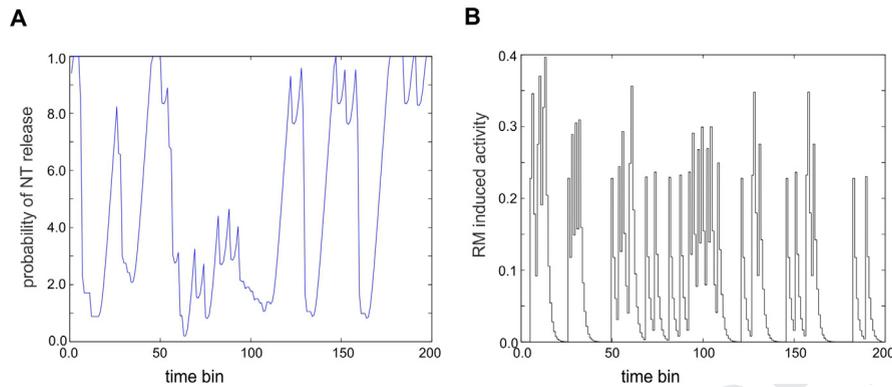


Fig. 4. Closed-loop based dynamics of neurotransmitter release probability and induced release machinery activity in a pre-synaptic neuron by diffused retrograde messenger from post-synaptic neuron at threshold (θ) equal to 1. The change in release probability as a result of increase in release machinery activity affects current into post-synaptic neurons. Consequently, the change of current into post-synaptic neurons alters release machinery activity that change release probability over time. For all simulations in this figure K and M values are used as 0.02 and 0.2, respectively. τ is set to 5 in the simulation.

142 0.2), K and M values are set to 0.2 and 0.1, respectively. Then the stimulation level is increased and the optimal values of θ
 143 and τ_{RM} are investigated to keep the L_3 firing rate at low levels (Fig. 9).

144 2.4. Computational complexity of the system

145 The proposed neural system incorporates a bio-inspired synaptic mechanism to show robustness to environmental
 146 changes that is modeled as different stimulation into L_1 neurons and is expressed as L_1 firing rate. It is expected that
 147 such simplified mechanisms do not increase running time of the algorithm remarkably when network size is increased. For
 148 this purpose, the running time of the algorithm in different network sizes and corresponding connection numbers are mea-
 149 sured (K and M values are set to 0.2 and 0.1, $\theta=4$ and $\tau_{RM}=50$) (Fig. 10). Further, the running time is measured for both
 150 the Integrated and Fire neuron and Izhikevich neuron model [41].

151 2.5. Simulations

152 The stimulation of the neural system from the environment is modeled as randomly selected 50% neurons in L_1 in each
 153 algorithm's run. Firing rate of L_1 neurons is modeled as a probabilistic approach described in the method. L_2 neurons are
 154 activated as Integrate and Fire neurons and their spiking patterns in each time bin are integrated and used to stimulate L_3
 155 neurons (motor neurons). L_3 neurons' activity is modeled as Integrate and Fire neurons and the average of their activities
 156 in each time bin is collected and the mean is presented. 500 times run of algorithm is used to calculate mean firing rate of
 157 L_3 neurons in all simulations. Connectivity rate of layers is also modeled as a probabilistic approach where each rate value
 158 is considered as probability of connecting of L_1 to L_2 or L_2 to L_3 . In each algorithm run (and fix connectivity rate) new
 159 connectivity matrix of layers are constructed. In all simulations 200 seconds stimulation of L_1 neurons is applied as time
 160 window time to collect data. In each second 100 time bins is considered to measure average neurons activity (spikes).

161 3. Results

162 The main aim of this study is to model a closed loop synaptic mechanism in postsynaptic neurons in a feedforward
 163 network that enables them to control neurotransmitter release from presynaptic neurons in response to different levels of
 164 input intensities. In our neural system, spiking activity of L_3 depends strongly on input from L_1 and the parameter values.
 165 Moreover, connectivity of layers plays an important role in transferring information from L_1 to L_3 . Therefore, it is important
 166 to study changes in firing rate of L_3 in different conditions and parameter values. The neurotransmitter release from L_2 to
 167 L_3 in the model depends on K and M values that determine change in probability of neurotransmitter release in each time
 168 bin and threshold of the release machinery (θ). Therefore, the dynamic of release machinery (RM induced activity) and its
 169 impact on probability of neurotransmitter release can be studied. Fig. 4A,B demonstrates the relationship between change
 170 of release probability and activity of release machinery in 200 time bins of stimulation of L_2 neurons for $\theta = 2$. In this
 171 simulation, K is set to 0.01 and M is set to 0.2. The threshold (θ) determines the tendency of pre-synaptic neuron to increase
 172 or decrease in release probability in each time bin. High threshold values leads to an increase in release probability over
 173 stimulation time.

174 To study the role of different K and M values in firing rate of L_3 neurons, threshold values (θ) equal to 1 to 4 are studied
 175 to measure firing rate of L_3 neurons (Fig. 5A-D). These simulations show that for all threshold values low K and high M
 176 values lead to higher firing rate of L_3 neurons. In addition, an increase in θ value results in an increase in firing rate of L_3
 177 neurons. In these simulations τ is set to 10 and firing rate of L_1 neurons is set to 0.9. In order to study the role of different

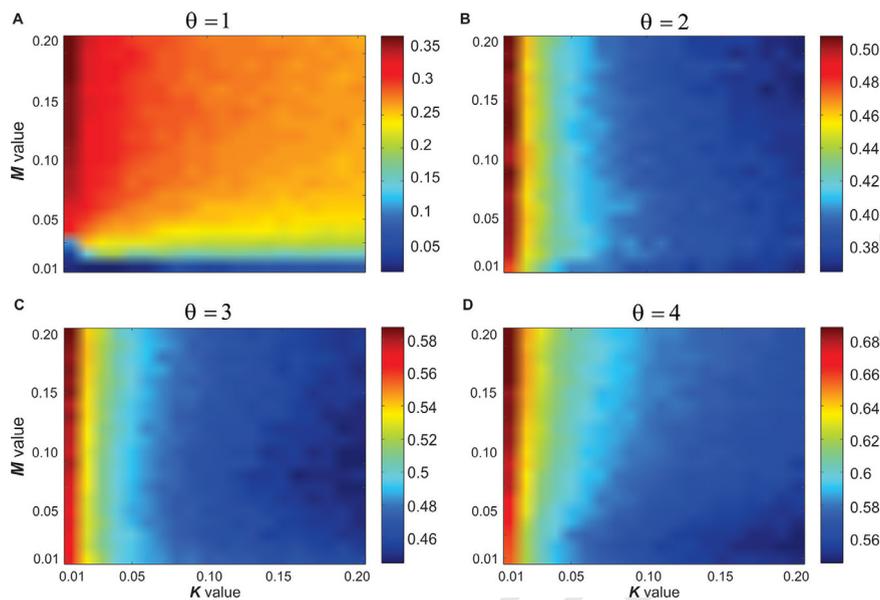


Fig. 5. The firing rate of motor neurons for different K and M parameter values and τ equal to 10. Figures **A** to **D** show firing rate for different K and M values and different thresholds of the controller. Higher threshold values cause a higher firing rate of motor neurons. The low threshold value decreases release probability and results in a decrease in current into motor neurons. In all conditions, lower K values and high M values lead to an increase in the average firing rate of motor neurons.

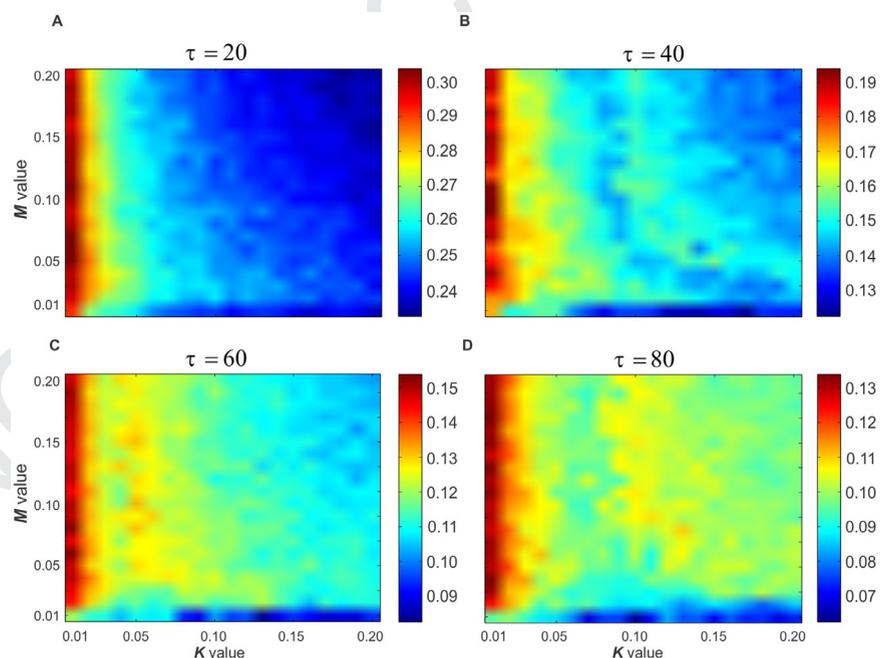


Fig. 6. Average firing rate of the motor neurons for different K and M parameter values for $\theta = 2$. To study the effect of incremental τ values on the firing rate of the motor neurons τ equal to 20, 40, 60 and 80 are used (**A** to **D**). The simulations show that an increase in τ value leads to decrease in firing rate of motor neurons for all pairs of K and M .

178 τ values in the process of firing rate of L_3 neurons, simulations are run for $\theta = 2$ and $\tau_{RM} = 10$ to 80. Simulations show
 179 decrease in K value and increase in M value lead to an increase in firing rate of L_3 neurons. Moreover, an increase in τ_{RM}
 180 values decreases the firing rate of L_3 neurons (Fig. 6A-D).

181 These results suggest that investigating the role of different θ and τ values in the firing rate of L_3 neurons. Fig. 7 shows
 182 the simulations for $K = 0.01$ and $M = 0.2$ values. Fig. 7A,B shows the results for connectivity between L_3 and L_2 equal to 0.4.
 183 Fig. 7C,D shows the results for connectivity between L_3 and L_2 equal to 0.9. As the firing rate of L_1 neurons may change

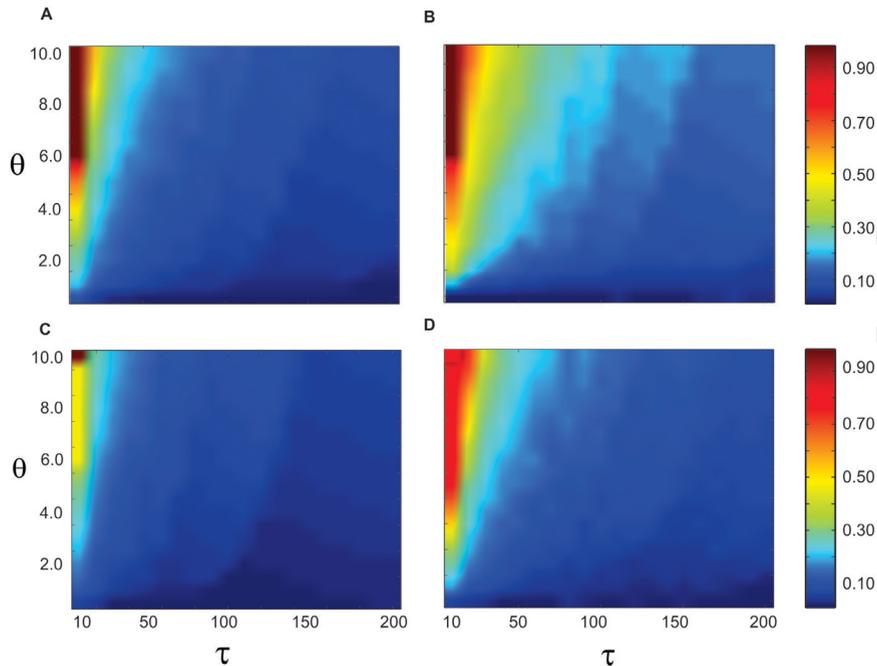


Fig. 7. Average firing rate of motor neurons for different θ and τ values. For each pair of θ and τ value, an average firing rate of the motor neurons for K equal to 0.01 and M equal to 0.2 is calculated. In figure **A** and **B** connectivity between second layer and the motor neurons is set to 0.9 and in figure **C** and **D** it is set to 0.2. The firing rate of the first layer is set to 0.4 (**A, C**) and 0.8 (**B, D**). In all simulations, lower τ and higher θ higher values increase the firing rate of the motor neurons.

184 in different stimulus presentation, different firing rate of L_1 neurons is also studied. For this purpose, $K=0.01$ and $M=0.2$
 185 values are used for different θ values are used in the simulation for connectivity rate between L_3 and L_2 equal to 0.9 (Fig. 8A)
 186 and equal to 0.2 (Fig. 8B). For each connectivity rate, average firing rate of a L_3 neuron (> 0) is calculated (Fig. 8C,D). These
 187 results show the possibility of controlling spiking activity of L_3 neurons by change in threshold value of L_2 neurons.

188 Fig. 9A,B shows the robustness of the system to different levels of stimulation to observe sparse and burst spiking by a
 189 change in θ and τ_{RM} . The results demonstrate that a gradual increase in τ_{RM} and decrease in θ leads to keep firing rate of L_1
 190 neurons at low levels (sparse spiking) when incremental levels of stimulations are presented to the system. To obtain burst
 191 spiking in decremented changes in stimulation level, θ values are decreased and τ_{RM} values are increased. Fig. 10A,B demon-
 192 strates the impact of network size and number of connections in the network on time of running the model (Fig. 10C,D).
 193 The results show that the model using Integrate and Fire model works faster than the model using Izhikevich neuron model.
 194 The results also show the impact of size on running time for both model neurons.

195 4. Discussion

196 Exploring the mechanisms that enable animals' neural systems to detect and encode stimuli in an environment plays an
 197 important role in understanding information processing in the brain.

198 Biological neurons are characterized by their different firing rate in response to stimuli according to stimulus intensity.
 199 Some neurons show burst spiking, while others show sparse spiking activity. Understanding the neural mechanism of spik-
 200 ing of different neurons in the brain plays a critical role in understanding the encoding and decoding mechanisms and
 201 applying them in artificial neural systems. It is likely that some neurons rely on some dendritic mechanisms like control-
 202 ling of neurotransmitter release by retrograde signalling to control the flow of activation signals from presynaptic neurons.
 203 However, neurons have no direct control on spiking activities of their inputs (the spiking of presynaptic neurons), though
 204 they may indirectly control the flow of information into their dendrites. Such mechanisms give the capability to control
 205 fluctuations in neuronal population activities triggered by stimuli in the environment. Specially, the role of retrograde sig-
 206 nals from postsynaptic to presynaptic neurons to change the probability of neurotransmitter release of presynaptic neurons
 207 is relatively well-known. Such changes in neurotransmitter release as a function of presynaptic neurons' activities can help
 208 prevent wasting neural metabolic energy to produce, release and uptake of neurotransmitters through neuronal communi-
 209 cation.

210 There are other simulation method that model neurotransmitter release based on spike timing [33], based on parameters
 211 influencing high calcium microdomains [3]. The role of interaction of fast retrograde signalling with Hebbian Plasticity to
 212 generate high encoding feedforward neural system has been theoretically studied [12]. In this work, a feedforward neural

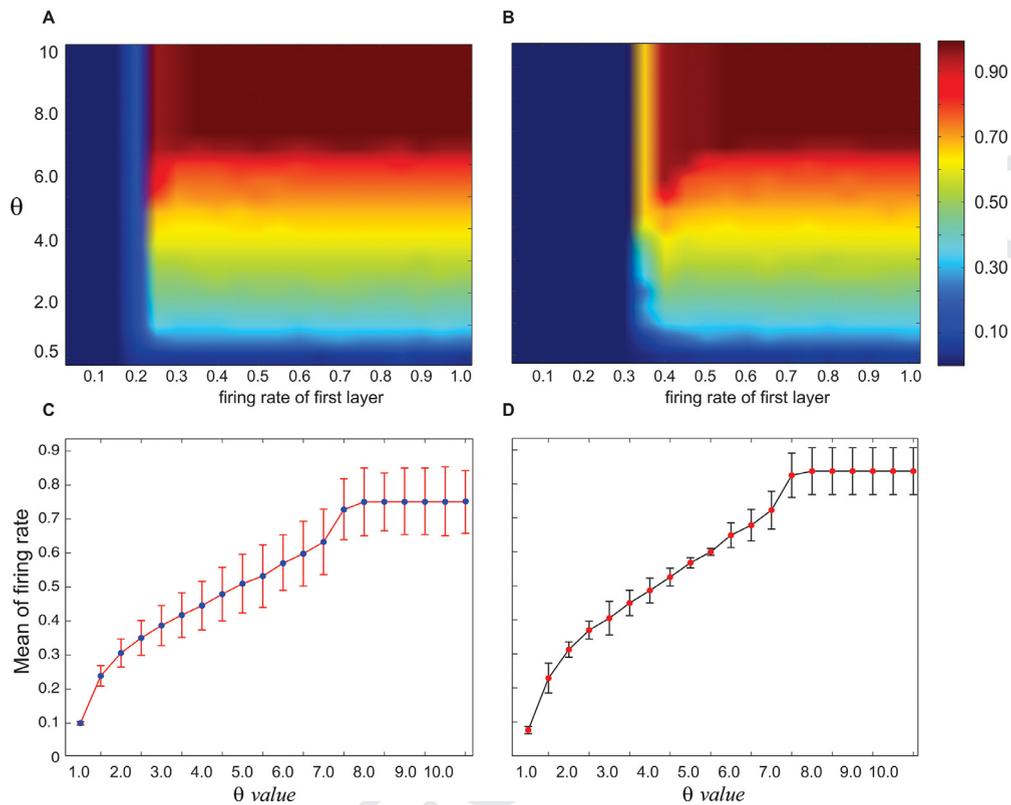


Fig. 8. The firing rate of the motor neurons vs. the firing rate of the first layer and different θ values at the connectivity rate between the second layer and the motor neurons are equal to 0.9 (A) and connectivity is equal to 0.3 (B). C. Mean of the firing rate of motor neurons vs. different θ values over firing rate of the first layer between 0.2 and 1.0 for connectivity is equal to 0.9 (C) and over 0.35 and 1.0 for connectivity is equal to 0.3 (D). The simulations show that different levels of motor neurons spiking rate the systems can be controlled by change in θ value. K value and M value used in the simulations is equal to 0.01 and 0.2, respectively. τ equal to 10 is used in all simulations.

213 network is presented that is able to detect stimuli. The synaptic mechanism that is modelled in the synapses between mo-
 214 tor neuron and presynaptic neurons enables the motor neuron to control the levels of input they receive over time. The
 215 controlling system acts as a closed-loop system that includes the production of different kinds of retrograde messengers in
 216 a postsynaptic neuron in response to influx current and their effect on neurotransmitter release from a presynaptic neuron.
 217 In our work, we presented a retrograde signaling based hypothetical mechanism for controlling neurotransmitter release in
 218 bio-inspired neural networks constrained by neurobiological data from insects' memory systems. Such changes in neuro-
 219 transmitter release influence the current into a postsynaptic neuron in each successive time bin. The amount of produced
 220 retrograde messenger in response to a given current in a time bin is determined by α parameter. In this study a moderate
 221 α value equal to 120 is used in all simulations. However, by changing α values, one can study the role of changes in firing
 222 rate of the neural system. Different retrograde messengers have a different rate of diffusion into presynaptic neurons and
 223 different speed of affecting the release machinery of the presynaptic neurons.

224 To our knowledge, this is the first work to model controlling of neurotransmitter release in sparse and burst spiking
 225 neurons robust to stimulation changes by a retrograde signalling based closed loop. The role of retrograde signaling from
 226 postsynaptic neurons into presynaptic neurons on release probability is known; however, the mechanisms of retrograde
 227 signalling's effect on cellular level and at different time scales are not fully known. Inspired by these observations, we
 228 developed a neural network model that can be applied in artificial systems including cognitive robotics where spiking neural
 229 networks are used to construct simple sensory-perception loops. Although variation in biophysical characteristics of neurons
 230 that induce sparse and burst spiking modes and their functional importance are under research, we assumed that retrograde
 231 signaling can play a critical role in controlling release probability (and so controlling functional spiking) when the neural
 232 system exist in dynamic environment with fluctuating stimulations. Moreover, this assumption and simplified controller
 233 developed in this study allows using a bio-inspired neural mechanism in artificial systems in the future.

234 In this work, in order to exhibit any spiking mode, electrical properties of presynaptic neurons are not affected. Instead,
 235 at the synaptic level, the probability of neurotransmitter release changes as a closed loop that modifies 'functional spiking
 236 of neurons', which is defined as spikes that trigger neurotransmitter release into the synaptic cleft. This work proposes
 237 novel experimental studies on the cellular mechanisms of neural adaptation to changes in stimulation of neurons in short
 238 term scale. This helps develop efficient artificial neural systems that exhibit different spiking patterns and able to shift from

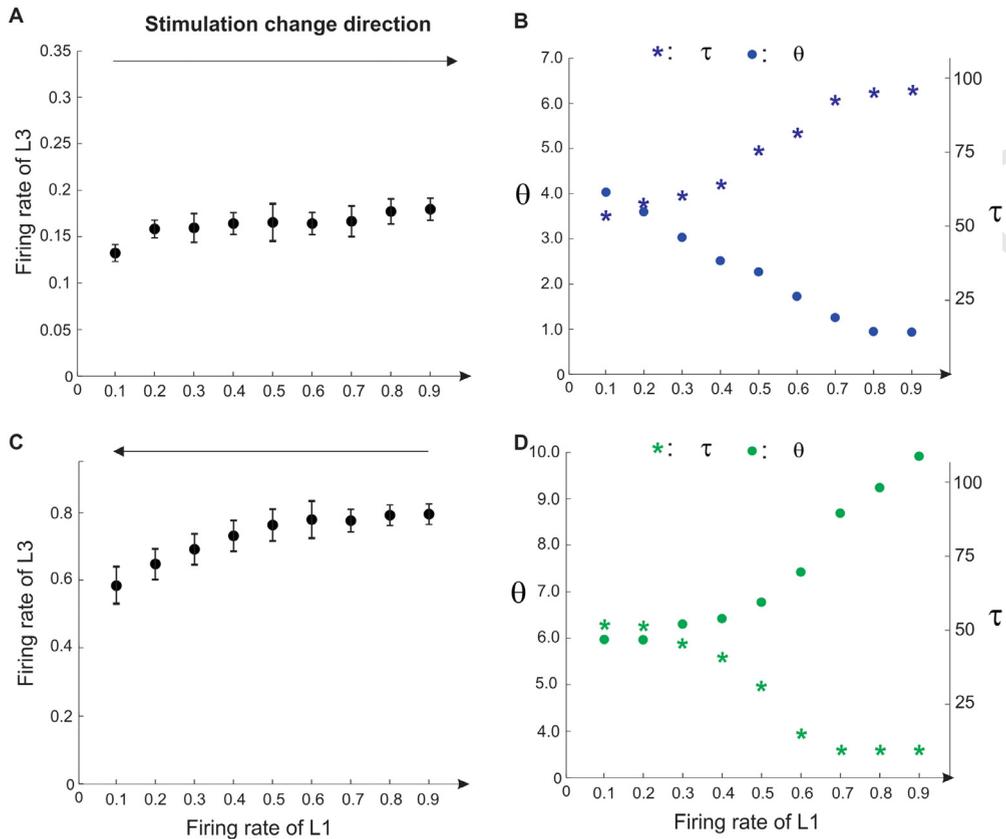


Fig. 9. Robustness of the systems in relation to changes in its stimulation by L_1 neurons (K and M value are set to 0.2). **A.** Sparse spiking activity of L_3 neurons by using optimal parameter conditions shown in **B.B.** Optimal parameter values to obtain minimum change in firing rate of sparse activity of L_3 neuron. At higher levels of stimulation (higher firing rate of L_1 neurons), τ should be increased while an increase in θ is required to keep L_1 firing rate at low values. For low stimulation as initial conditions, τ and θ values are set to 50 and 4, respectively. **C.** Burst activity of L_3 neurons for optimal parameter values shown in **D.** **D.** Optimal parameter values to obtain minimum change in burst activity of L_3 neuron. At lower levels of stimulation (lower firing rate of L_1 neurons), τ should be increased while a decrease in θ is required to keep L_1 firing rate at high values when stimulation is lowered. For high stimulation as initial conditions, τ and θ values are set to 10 and 10, respectively.

239 a spiking to a different mode. This simply occurs due to a change in two model parameters τ_{RM} and θ . Therefore, this
 240 synaptic closed loop allows for the development of Spiking Neural Networks with self control capabilities to be used in
 241 artificial systems including robotics.

242 The model exerts the biophysical features of retrograde messengers as different levels of τ_{RM} values. The results show
 243 that low τ_{RM} values leads to a higher firing rate of motor neurons due to a fast effect as well as a fast decay. But high τ_{RM}
 244 values lower firing rate as a consequence of accumulation of induced activity by retrograde messengers in pre-synaptic neu-
 245 rons. However, the neural system may benefit from high τ_{RM} values for long-term memory where the system needs to keep
 246 its high or low firing rate over a long time. Another parameter of the model that is involved in information processing of the
 247 developed neural system is the threshold of the release machinery (θ). In our model, an increase in threshold value leads
 248 to an increase in motor neuron's firing rate for a given M and K values and τ_{RM} value equal to 10. In general, high M and
 249 low K values result in higher motor neuron's firing rate. In the simulations, fixing threshold value and increasing τ_{RM} value
 250 resulted in a decrease of motor neurons firing rate. As the neural system showed robustness to fluctuation of firing rate of
 251 first layer neurons for a given θ value, Fig. 8 proposes a controlling mechanism of motor neurons' firing rate by changing θ
 252 values. If such a neural system is expected to show sparse spiking, it exerts lower θ values while high θ values equipped
 253 the neural system with burst spiking activity.

254 The theory that is presented here is on the mechanism of homeostatic regulation of neurotransmitter release as a prob-
 255 abilistic event by postsynaptic dendrites. It is based on hypothetical protein machinery or biochemical pathway that acts
 256 according to its activity level compared to its threshold of shifting increases or decreases in release probability. This study
 257 assigns an important role for a molecular mechanism in the neurons that are able to help modify synaptic information flow.
 258 Therefore, it proposes experimental investigations to test such hypothesis involved in dendritic computations.

259 Theoretical studies can help understand neuronal computations using novel models and simulations that can also be used
 260 in developing artificial spiking neural systems. One of the challenges in developing next generation of artificial systems (e.g.

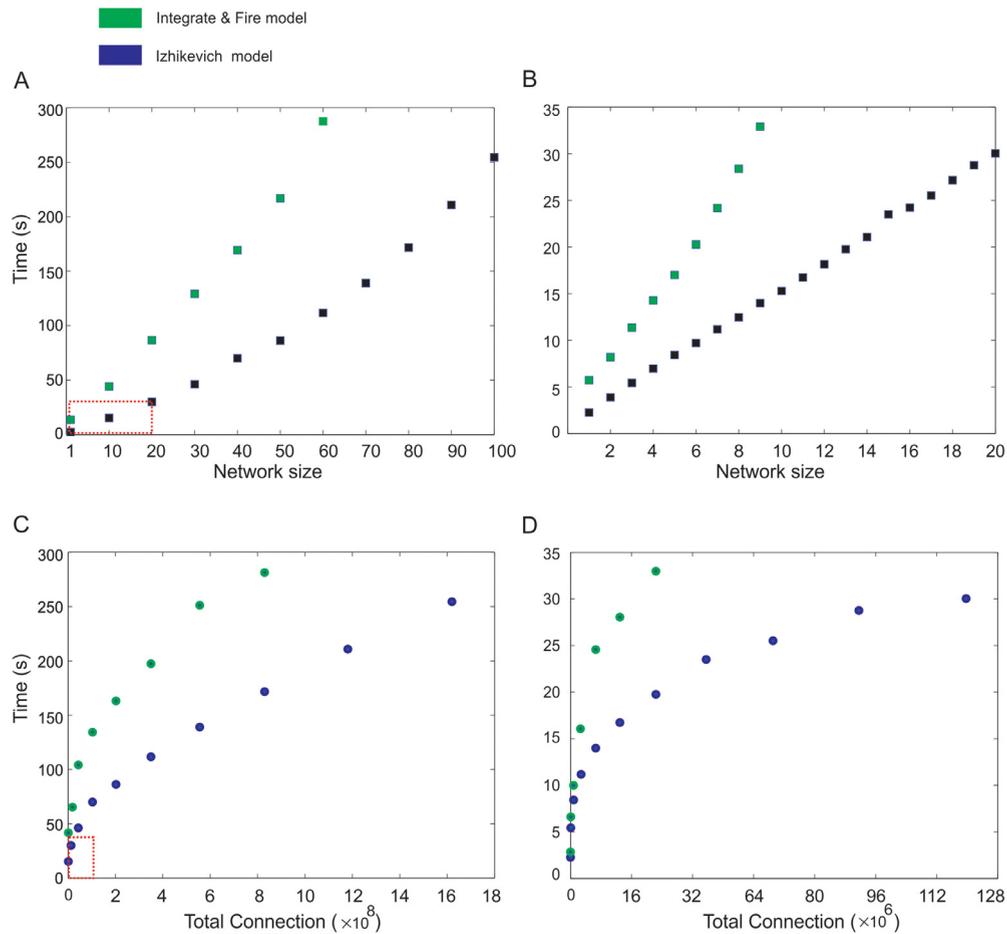


Fig. 10. Numerical results on the computational complexity of the model to study the impact of neuron model used in simulations. Integrate and fire vs. Izhikevich models are compared. The time of running algorithm for a given set of parameter values versus network size and network connections are shown in (A, B) and (C, D), respectively. Initial size of the network in all simulations is 10, 60 and 10 neurons in L_1 , L_2 , and L_3 , respectively. The connectivity rate of L_1 - L_2 and L_2 - L_3 is set to 0.9 and 0.3, respectively. **A.** Network size is between 1 and 100 times initial size of the network and between 1 and 20 in **B.** Time of running program is measured versus total number of connection of layers for whole network size (C) and network size between 1 and 20 (D). The results show that compared to the Izhikevich model, the Integrate and Fire model can lead to a lower running time of the algorithm.

261 navigating robots) is how to implement the efficiency of animals' brain in encoding complex environments enriched with
 262 different kinds of stimuli that should be detected, encoded and stored in the neural networks [37]. Therefore, one fun-
 263 damental step to generate new artificial systems would be to understand basic information processing in neural circuits
 264 underlying behavior (e.g. learning tasks). This includes understanding molecular, cellular, synaptic and network level mech-
 265 anisms. This study is aimed to illustrate the potential of developing bio-inspired neural systems that are equipped with
 266 simplified synaptic communication in biological neurons. One challenge for developing a class of spiking neural networks is
 267 how to implement a simplified model of complicated cellular mechanisms in biological neurons. For this purpose, we pro-
 268 pose to investigate the possibility of new classes of bio-inspired neural networks that are highly similar to biological neural
 269 systems.

270 The structural and physiological parameters in biological neural systems determine their capability to exhibit cognitive
 271 functions like learning and memory. The existence of different mechanisms of information processing strongly depends on
 272 the brain region and neuron type. Combining different mechanisms at different levels (molecular, synaptic, cellular and
 273 network) across different time scales of an event into models, may play an important role in exploring neural circuits of
 274 different cognitive capabilities of animals' brains. For example, neural systems may gain benefits from STDP and Hebbian
 275 mechanisms in combination with retrograde signalling based mechanisms. However, it is necessary to develop simulations
 276 using integrated mechanisms in different paradigms to explore the importance of such combined mechanisms. The work
 277 presented here can enhance our understanding of complex strategies that have been developed through evolution and used
 278 by the human and animal brain for information processing and intact behaviour.

279 5. Conclusion

280 Brain-like artificial architectures using spiking neural networks (SNN) have many industrial applications including cogni-
 281 tive robotics [39]. To achieve this goal the main challenge is to understand morphological and electrophysiological variations
 282 observed in biological neural networks. Studying the cellular and synaptic mechanisms and function of sparse and burst
 283 neuronal activities play critical role in understanding how information is transformed in brains.

284 By exploring the role of neuronal architecture and information processing of different neural systems, biologically plau-
 285 sible brain-like artificial systems can be developed in future.

286 In this work some synaptic and cellular and network knowledge on biological systems are used to develop a feedforward
 287 neural system that is capable to control the functional activity of its neurons in different levels of environmental stimulation.
 288 Simplified mechanisms implemented in this work allow industrial application of the model and propose some possible
 289 mechanisms in neurons to investigate as well.

290 This work proposes novel experimental studies on the role of retrograde signaling in short time scale on the controlling
 291 of neurotransmitter release of presynaptic neurons by postsynaptic neurons. This work presents a cellular hypothesis on
 292 how neurons exhibit persistent sparse or burst spiking activity by changing their probability of neurotransmitter release in
 293 synapses.

294 References

- 295 [1] O. Akyol, et al., Nitric oxide as a physiopathological factor in neuropsychiatric disorders, *In vivo* 18 (3) (2004) 377–390.
 296 [2] L.F. Abbott, G.R. Wade, Synaptic computation, *Nature* 431 (7010) (2004) 796–803.
 297 [3] R. Bertram, G.D. Smith, A. Sherman, Modeling study of the effects of overlapping Ca²⁺ microdomains on neurotransmitter release, *Biophys. J.* 76 (2)
 298 (1999) 735–750.
 299 [4] T.V.P. Bliss, G.L. Collingridge, R.G.M. Morris, Synaptic plasticity in health and disease: introduction and overview, *Philos. Trans. R. Soc. B: Biol. Sci.* 369
 300 (1633) (2014) 20130129.
 301 [5] C.S. Bingham, et al., A large-scale detailed neuronal model of electrical stimulation of the dentate gyrus and perforant path as a platform for electrode
 302 design and optimization, in: *Engineering in Medicine and Biology Society (EMBC), 2016 IEEE 38th Annual International Conference of the, IEEE, 2016,*
 303 pp. 2794–2797.
 304 [6] F. Cao, D. Wang, H. Zhu, Y. Wang, An iterative learning algorithm for feedforward neural networks with random weights, *Inf. Sci.* 328 (2016) 546–557.
 305 [7] H.K. Chan, D.P. Yang, C. Zhou, T. Nowotny, Burst firing enhances neural output correlation, *Front. Comput. Neurosci.* 10 (2016).
 306 [8] I. Elices, P. Varona, Asymmetry factors shaping regular and irregular bursting rhythms in central pattern generators, *Front. Comput. Neurosci.* 11 (2017).
 307 [9] F. Faghihi, A.A. Moustafa, The dependence of neuronal encoding efficiency on Hebbian plasticity and homeostatic regulation of neurotransmitter re-
 308 lease, *Front. Cell. Neurosci.* 9 (2015).
 309 [10] F. Faghihi, A.A. Moustafa, Impaired homeostatic regulation of feedback inhibition associated with system deficiency to detect fluctuation in stimulus
 310 intensity: a simulation study, *Neurocomputing* 151 (2015) 1248–1254.
 311 [11] F. Faghihi, A.A. Moustafa, A computational model of pattern separation efficiency in the dentate gyrus with implications in schizophrenia, *Front. Syst.*
 312 *Neurosci.* 9 (2015) 42.
 313 [12] F. Faghihi, A.A. Moustafa, The dependence of neuronal encoding efficiency on Hebbian plasticity and homeostatic regulation of neurotransmitter re-
 314 lease, *Front. Cell. Neurosci.* 9 (2015).
 315 [13] S.M. Farris, Are mushroom bodies cerebellum-like structures? *Arthropod Struct. Dev.* 40 (4) (2011) 368–379.
 316 [14] M. Gómez-Gonzalo, et al., Endocannabinoids induce lateral long-term potentiation of transmitter release by stimulation of gliotransmission, *Cerebral*
 317 *Cortex* (2014) bhu231.
 318 [15] A.W. Harrington, D.D. Ginty, Long-distance retrograde neurotrophic factor signalling in neurons, *Nat. Rev. Neurosci.* 14 (3) (2013) 177–187.
 319 [16] M. Heisenberg, What do the mushroom bodies do for the insect brain? An introduction, *Learn. Memory* 5 (1) (1998) 1–10.
 320 [17] C. Hong, J. Yu, J. Wan, D. Tao, M. Wang, Multimodal deep autoencoder for human pose recovery, *IEEE Trans. Image Process.* 24 (12) (2015) 5659–5670.
 321 [18] C. Hong, J. Yu, D. Tao, M. Wang, Image-based three-dimensional human pose recovery by multiview locality-sensitive sparse retrieval, *IEEE Trans. Ind.*
 322 *Electron.* 62 (6) (2015) 3742–3751.
 323 [19] T. Iliou, C.N. Anagnostopoulos, I.M. Stephanakis, G. Anastassopoulos, A novel data preprocessing method for boosting neural network performance: a
 324 case study in osteoporosis prediction, *Inf. Sci.* 380 (2017) 92–100.
 325 [20] E.M. Izhikevich, Simple model of spiking neurons, *IEEE Trans. Neural Netw.* 14 (6) (2003) 1569–1572.
 326 [21] E.M. Izhikevich, Which model to use for cortical spiking neurons? *IEEE Trans. Neural Netw.* 15 (5) (2004) 1063–1070.
 327 [22] P. Jia, J. Yin, D. Hu, Z. Zhou, Retrograde adaptive resonance theory based on the role of nitric oxide in long-term potentiation, *J. Comput. Neurosci.* 23
 328 (1) (2007) 129–141.
 329 [23] P.D. King, J. Zylberberg, M.R. DeWeese, Inhibitory interneurons decorrelate excitatory cells to drive sparse code formation in a spiking model of V1, *J.*
 330 *Neurosci.* 33 (13) (2013) 5475–5485.
 331 [24] M.G. Metzner, R. Krahe, M.J. Chacron, Burst firing in the electrosensory system of gymnoti form weakly electric fish: mechanisms and functional roles,
 332 *Front. Comput. Neurosci.* 10 (2016).
 333 [25] Ohno-Shosaku, T., Kano, M., Endocannabinoid-mediated retrograde modulation of synaptic transmission, *Curr. Opin. Neurobiol.*, 29, (2014) 1–8.
 334 [26] K.E. Perks, T.Q. Gentner, Subthreshold membrane responses underlying sparse spiking to natural vocal signals in auditory cortex, *Eur. J. Neurosci.* 41
 335 (5) (2015) 725–733.
 336 [27] M. Richert, J.M. Nageswaran, N. Dutt, J.L. Krichmar, An efficient simulation environment for modeling large-scale cortical processing, *Front. Neuroinf.*
 337 (5) (2011).
 338 [28] J.R. Steinert, T. Chernova, I.D. Forsythe, Nitric oxide signaling in brain function, dysfunction, and dementia, *Neuroscientist* 16 (4) (2010) 435–452.
 339 [29] A. Salehi, J.-D. Delcroix, W.C. Mobley, Traffic at the intersection of neurotrophic factor signaling and neurodegeneration, *Trends Neurosci.* 26 (2) (2003)
 340 73–80.
 341 [30] Y. Suvarna, N. Maity, M.C. Shivamurthy, Emerging trends in retrograde signaling, *Mol. Neurobiol.* 53 (4) (2016) 2572–2578.
 342 [31] B. Schmidt, D.F. Marrone, E.J. Markus, Disambiguating the similar: the dentate gyrus and pattern separation, *Behav. Brain Res.* 226 (1) (2012) 56–65.
 343 [32] D.M. Schneider, S.M. Woolley, Sparse and background-invariant coding of vocalizations in auditory scenes, *Neuron* 79 (1) (2013) 141–152.
 344 [33] W. Senn, H. Markram, M. Tsodyks, An algorithm for modifying neurotransmitter release probability based on pre- and postsynaptic spike timing, *Neural*
 345 *Comput.* 13 (1) (2001) 35.
 346 [34] G. Turrigiano, Too many cooks? Intrinsic and synaptic homeostatic mechanisms in cortical circuit refinement, *Ann. Rev. Neurosci.* 34 (2011) 89–103.
 347 [35] H.W. Tao, M.M. Poo, Retrograde signaling at central synapses, *Proc. Natl. Acad. Sci.* 98 (20) (2001) 11009–11015.
 348 [36] Tetzlaff, C., Kolodziejcki, C., Markelic, I., Wörgötter, F., Time scales of memory, learning, and plasticity, *Biol. Cybern.*, 106(11–12), (2012) 715–726.
 349 [37] J. Triesch, Synergies between intrinsic and synaptic plasticity mechanisms, *Neural Comput.* 19 (4) (2007) 885–909.

- 350 [38] P.J. Uhlhaas, W. Singer, Oscillations and neuronal dynamics in schizophrenia: the search for basic symptoms and translational opportunities, *Biol.*
351 *Psychiatry* (2014).
- 352 [39] Y. Wang, Cognitive robotics and mathematical engineering, *Cognitive Informatics & Cognitive Computing (ICCI* CC)*, 2015 IEEE 14th International
353 Conference on, IEEE, 2015.
- 354 [40] J. Wolfe, A.R. Houweling, M. Brecht, Sparse and powerful cortical spikes, *Current Opin. Neurobiol.* 20 (3) (2010) 306–312.
- 355 [41] J. Wang, Q. Cai, Q. Chang, J.M. Zurada, Convergence analyses on sparse feedforward neural networks via group lasso regularization, *Infor. Sci.* 381
356 (2017) 250–269.
- 357 [42] D.G. Wüstenberg, M. Boytcheva, B. Grünewald, J.H. Byrne, R. Menzel, D.A. Baxter, Current and voltage-clamp recordings and computer simulations of
358 Kenyon cells in honeybees, *J. Neurophysiol.* 92 (4) (2004) 2589–2603.
- 359 [43] J. Yu, X. Yang, F. Gao, D. Tao, Deep multimodal distance metric learning using click constraints for image ranking, *IEEE Trans. Cybern* (2016).
- 360 [44] J. Yu, B. Zhang, Z. Kuang, D. Lin, J. Fan, iPrivacy: image privacy protection by identifying sensitive objects via deep multi-task learning, *IEEE Trans. Inf.*
361 *Forensics Secur.* 12 (5) (2017) 1005–1016.
- 362 [45] F. Zenke, E.J. Agnes, W. Gerstner, Diverse synaptic plasticity mechanisms orchestrated to form and retrieve memories in spiking neural networks,
363 *Nature Commun.* (6) (2015).